Total phenolic compounds, antioxidant activity and toxicity of selected medicinal and aromatic plants

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etary antioxidant components for the prevention of some diseases and health quality improvement has attracted much research attention through the last decade. Vegetables and herbal influsions d as important antioxidant sources. Food industry shows significant interest in application of plant bioactive compounds for flavoring but also for preservation purposes, but attention should be given in In the current study were investigated the total phenolic content, the antioxidant activity and the toxicity of selected medicinal aromatic plants that are being consumed as decoctions or used as food

2 g dry mass of each plant species were:

ed in 200 ml (1 cup) a) at 85 °C for 15 min,

with the assistance of ultrasonic

for 15 min and.

further extracted by petroleum

The herbal infusions were then nan filter No.1.

nt (in terms of caffeic acid) was determined using a Folin-Ciocalteu assay. Antioxidan ooth the ABTS assay - 2,2-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid), and the DPPH oicrylhydrazyl radical .

t of the plant extracts has been performed using MICROTOX® toxicity analyzer. MICROTOX® minescence bacteria Vibrio fischeri as a reference test species for the measurement of ition. The protocol used was the Basic Test 81.9% (AZUR, 1997) with appropriate primary order the EC_{50} (Effective Concentration where bioluminescence inhibition is 50%) not to be

Code				
MOF	Melissa officinalis	Lemon balm	leaves	Decoctions
OVU	Origanum vulgare	Oregano	leaves	Herb in salads (fresh or dry), roasted, stewed
ODI	Origanum dictamnus	Dittany	Leaves/flowers	Decoctions
SOF	Salvia officinalis	Sage	Leaves	Decoctions, as a herb in cooking
HOF	Hyssopus officinalis	Hyssop	Leaves/flowers	Decoctions

Table 2. Toxicity of pla	ant species expressed as	es expressed as EC50 (mg/ml) after 15 minutes incubation time			
			Treat	men	
	(a)	(a-pte)*1	(b)		

	(a)	(a-pte)*1	(b)	(b-pte)*1	(c)	(c-pte)*1
MOF	119.75	210.68	200.61	216.30	113.09	126.93
HOF	38.31	139.60	150.67	162.75	96.05	101.47
ODI	10.56	42.87	60.46	110.75	62.61	96.87
ovu	8.61	36.38	40.96	52.32	22.09	77.69
SOF	52.17	107.77	199.45	198.78	137.19	224.87

MOF Melissa officinalis		197.0	1268.3	1321.7
HOF Hyssopus officinalis		34.9	206.4	206.8
OVU Origanum vulgare	(a)	128.6	631.1	669.3
ODI Origanum dictamnus		63.9	299.7	321.0
SOF Salvia officinalis		64.5	328.2	326.1
MOF Melissa officinalis		185.9	1240.1	1319.5
HOF Hyssopus officinalis		31.7	182.5	190.2
OVU Origanum vulgare	(a – pte)	105.1	565.2	595.2
ODI Origanum dictamnus		51.3	227.7	311.9
SOF Salvia officinalis		62.0	306.9	323.1
MOF Melissa officinalis		135.8	644.5	766.8
HOF Hyssopus officinalis		10.7	51.5	69.2
OVU Origanum vulgare	(b)	63.5	290.1	309.5
ODI Origanum dictamnus		28.3	153.5	169.6
SOF Salvia officinalis		15.5	77.3	99.2
MOF Melissa officinalis		119.1	605.5	684.3
HOF Hyssopus officinalis		10.4	39.2	71.0
OVU Origanum vulgare	(b – pte)	53.9	262.8	267.1
ODI Origanum dictamnus		27.9	123.7	139.1
SOF Salvia officinalis		15.5	66.3	89.6
MOF Melissa officinalis		159.9	752.7	871.8
HOF Hyssopus officinalis		13.1	49.7	85.2
OVU Origanum vulgare	(c)	68.3	323.5	344.5
ODI Origanum dictamnus		34.5	177.0	179.6
SOF Salvia officinalis		16.9	83.7	123.1
MOF Melissa officinalis		114.7	676.0	834.9
HOF Hyssopus officinalis		15.0	55.7	86.1
OVU Origanum vulgare	(c - pte)	57.6	276.3	288.7
ODI Origanum dictamnus		32.9	155.5	160.3
SOF Salvia officinalis		28.0	68.6	107.9

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Code					
MOF	(b)	92.33	18.47	200	
MOF	(a)	48.00	9.60	200	Decoction
	(b)	12.95	6.48	500	In salads, appetizers and meals
OVU	(a)	3.29	0.66	200	Decoction
	(a)		16.5	5000	During cooking process
0.01	(b)	21.03	4.21	200	
ODI	(a)	4.04	0.81	200	Decoction
	(b)	83.79	16.76	200	
SOF	(a)	18.95	3.79	200	Decoction
	(a)		94.75	5000	During cooking process
HOF	(b)	65.10	13.02	200	
HOF	(a)	11.90	2.38	200	Decoctions

Results

Total phenolic content was estimated in both infusions and the aqueous phase of their extether, expressed in mg of caffeic acid /200 ml. Maximum values were given by Melissa officia antioxidalt activity regardless of the method that is being determined DPPH or ABTS is ranking of the total phenolic content. Results show that there is a scaling increase in to among the species tested. Melissa officinalis exhibited the higher phenolic content and Hys lowest regardless the extraction procedure. Ranking remains the same in the aquatic phase petroleum ether for all the infusions (table 2) tested.

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Antioxidant activity when determined by ABTS method expressed also umole Trolox / 200 m Maximum values were given by Melissa officinalis. The ranking of antioxidant activity rega that is being determined DPPH or ABTS is the same with the ranking of the total phenolic co

that is being determined DPPH or ABTS is the same with the training of the state of the plant extracts was performed by applying the Basic Test protocol of exposure expressed as $\mathrm{EC_{20}}$ value, at which 50% loss of luminescence is obtained, $\mathrm{EC_{20}}$ wa which a 20% loss of light emission is observed, as according to the ISO guidelines (ISO 1 sample shows an effect percentage greater than 20%. Results show that $Organum\ vulgare\ values$ in all treatments. Respectively, higher values were obtained by $Melissa\ officinalis\ in\ pte$, (b), (b-pte), and by $Salvia\ officinalis\ in\ treatments\ (c)\ and\ (c-pte)\ (table\ 3).$ Depending on the use of each plant tested in nutrition, was calculated the maximum dry plant decoctions, in 500ml in case of use in salads, and in full when is being used in cooking proce Furthermore, was calculated the synergism ratio (SR) as follows: $SR_{total\ phenotics} = Total\ phenotics\ - pte\ /\ Total\ phenotic mixture$ $SR_{unitoordant\ activity} = Antioxidant\ activity - pte\ /\ Antioxidant\ activity\ mixture$ $SR_{total\ phenotics} = SR_{total\ pte} = CS_{50}\ pte\ /\ ECS_{50}\ mixture$

Sample Code	Treatment	SR total phenolic	Effect *1	SR DPPH	Effect *1	SR ABTS	Effect *1	SR toxicity*2	Effect
	(b)	0.9	Synergism	0.9	Synergism	0.9	Synergism	1.1	Synergism
MOF	(c)	0.7	Synergism	0.9	Synergism	1.0	Additive	1.1	Synergism
	(a)	0.9	Synergism	1.0	Additive	1.0	Additive	1.8	Synergism
	(b)	1.0	Additive	0.8	Synergism	1.0	Additive	1.1	Synergism
HOF	(c)	1.1	Antagonism	1.1	Antagonism	1.0	Additive	1.1	Synergism
	(a)	0.9	Synergism	0.9	Synergism	0.9	Synergism	3.6	Synergism
	(b)	1.0	Additive	0.8	Synergism	0.8	Synergism	1.8	Synergism
ODI	(c)	1.0	Additive	0.9	Synergism	0.9	Synergism	1.5	Synergism
	(a)	0.8	Synergism	0.8	Synergism	1.0	Additive	4.1	Synergism
ovu	(b)	0.8	Synergism	0.9	Synergism	0.9	Synergism	1.3	Synergism
	(c)	0.8	Synergism	0.9	Synergism	0.8	Synergism	3.5	Synergism
	(a)	0.8	Synergism	0.9	Synergism	0.9	Synergism	4.2	Synergism
SOF	(b)	1.0	Additive	0.9	Synergism	0.9	Synergism	1.0	Additive
	(c)	1.7	Antagonism	0.8	Synergism	0.9	Synergism	1.6	Synergism
	(a)	1.0	Additive	0.9	Synergism	1.0	Additive	2.1	Synergism

Conclusions. Results of this study indicate that between plant species examined, Melissa of the higher, and Hyssopus officinalis the lower phenolic content and actioxidant activity. E seems to influence significantly the extraction of phenolic compounds and antioxidant activ great differences were observed between extractions in high and room temperature. A observed in ultrasound assisted extraction. Similar results were obtained when infu petroleum ether. Linear positive correlation has been found between total phenolic contractivity in the infusions studied.

Toxicity of infusions seems to be also influenced by the temperature and extraction correlation was detected between toxicity and the total phenolic content or antioxidant act soluble substances and the essential oil of plants were remarkable in case of toxicity in cor total phenolic content and antioxidant activity where interaction was almost neutral

tary plants. Food and Chemical Toxicology 46, 3325-3332. ACHN and A2780/CP cell lines. Computers in Biology and Medicine 41, 665-674. hemosphere 68, 724-730.











